



Mahajana Education Society (R)

Education to Excel

SBRR Mahajana First Grade College (Autonomous)

Post Graduate Wing

Pooja Bhagavat Memorial Mahajana Education Centre

KRS road, Metagalli, Mysuru

Department of Studies in Biotechnology

Credit Based Choice Based Continuous Evaluation Pattern System

M.Sc. Biotechnology

Eligibility for admission: Candidates with Bachelor's Degree in Science including Agricultural, Pharmacy, Chemical Engineering, Medicine, Veterinary, Dairy, Fisheries, Horticulture, Forestry from any University recognized by UGC / ICAR / AICTE / Medical Council with an aggregate minimum of 55% (50% in case of SC/ST) or equivalent grade.

Scheme & Credit pattern
Credit Based Choice Based Continuous Evaluation Pattern System
SCHEME OF THE STUDY

MINIMUM CREDITS TO BE REGISTERED BY A STUDENT IN A NORMAL PHASE TO SUCCESSFULLY COMPLETE M.Sc. BIOTECHNOLOGY DEGREE IN FOUR SEMESTERS

Semesters	Hardcore		Softcore		Open elective		Total	
	Numbers	Credits	Numbers	Credits	Numbers	Credits	Numbers	Credits
I semester	04	14	02	06	-	-	06	20
II semester	04	14	02	06	01	04	07	24
III semester	04	14	02	06	-	-	06	20
IV semester	01	09	01	03	-	-	02	12
Total	13	51	09	21	01	04	21	76

Course Type	Credits
Hard Core	Minimum credits 42 and maximum credits 52
Soft Core	Minimum credits 16
Open Elective	Minimum credits 04
Total	76

Semester-I Credits: 20

NO	PAPER CODE	TITLE OF THE COURSE PAPER	CREDITS
1	HC	Bioanalytical Techniques	3
2	HC	Microbiology	3
3	HC	Biochemistry	3
4	HC	Practical-1 (Bioanalytical Techniques, Microbiology, Biochemistry)	5
		Select 2 among 3	
5	SC	Cancer Biology	3
6	SC	Food & Environmental Biotechnology	3
7	SC	Biostatistics & Bioinformatics	3

Semester-II Credits: 24

NO	PAPER CODE	TITLE OF THE COURSE PAPER	CREDITS
1	HC	Molecular Biology	3
2	HC	Genetic Engineering	3
3	HC	Immunotechnology	3
4	HC	Practical-2 (Molecular Biology, Genetic Engineering, Immunotechnology)	5
		Select 2 among 3	
6	SC	Molecular Genetics	3
7	SC	Genomics & Proteomics	3
8	SC	Cell Biology and Cellular Signalling	3
9	OE	Biotechnology and its applications (For other discipline students)	3

Semester-III Credits: 20

NO	PAPER CODE	TITLEOF THE COURSE PAPER	CREDITS
1	HC	Plant Biotechnology	3
2	HC	Animal Biotechnology	3
3	HC	Bioprocess Technology	3
4	HC	Practical-3 (Plant and Animal Biotechnology, Bioprocess Technology)	5
		Select 2 among 3	
	SC	Molecular Diagnostics	3
4	SC	Natural Products &Drug Discovery	3
5	SC	Nanobiotechnology	3
6	SC	Molecular Diagnostics	3

Semester-IV Credits: 12

NO	PAPER CODE	TITLEOF THE COURSE PAPER	CREDITS
1	SC	Stem Cell & Regenerative Medicine	3
2	SC	Molecular Plant Pathology	3
3	HC	Project work/Dissertation*	9

Additional Softcores

NO	PAPER CODE	TITLEOF THE COURSE PAPER	CREDITS
1	SC	Bioentrepreneurship	3
2	SC	Seed Health and Diagnostics	3

I semester
Bioanalytical techniques (HC)

48 h

Unit I

General considerations, pH and buffers, cell disruption techniques. Cell fractionation, Lysis buffer, Salting in and salting out, dialysis.

Chromatographic techniques: General principles, Sample preparation, Selection of chromatographic system, Low pressure column chromatography, HPLC, Adsorption chromatography, Partition chromatography, Ion exchange chromatography, Exclusion chromatography, Affinity chromatography, GLC, TLC, Paper chromatography. UPLC, chromatofocusing

Unit-II

Electrophoretic Techniques: General principles, Support media, Native gels, SDS-PAGE, IEF, 2D gel electrophoresis, Agarose gel electrophoresis, Pulse field gel electrophoresis (PFGE), Capillary electrophoresis (CE).

Centrifugation Techniques: Introduction, Basic principles of sedimentation, Types of centrifuges and their uses, Preparative centrifugation- differential and density gradient separation, Analytical ultracentrifuges and their applications.

Zymogram, reverse zymogram,, Visualising the separated components, staining, fluorescence, PAS staining.

Unit-III

Spectroscopic techniques: Introduction, UV and visible light spectroscopy, IR and Raman spectroscopy, Electron Spin Resonance (ESR), NMR, Spectrofluorimetry, Luminometry, Atomic spectroscopy, X-ray diffraction, ORD, CD.

Mass spectrometric techniques: Introduction, mass spectrometer, Ionization techniques- Electron impact ionization (EI), Electrospray Ionization, Chemical ionization (CI), Field ionization (FI), MALDI, Ion disruption methods, Ion desorption and evaporation methods, Analyzers- Magnetic sector, time-of-flight, quadrupole, ion trap, Detectors- electron multipliers, Tandem mass spectrometry, applications.

Unit-IV

Microscopy techniques: Light microscopy, phase contrast microscopy, fluorescence microscopy, electron microscope- TEM and SEM, confocal microscopy, flow cytometry- FACS.

Radioisotope techniques: Nature of radioactivity, detection and measurement, GM counter, scintillation counting, autoradiography, Safety aspects and applications of radioisotopes in biology.

Electrochemical techniques: Introduction, Principles, Redox reactions, Types of electrodes- pH electrode, ion-sensing electrodes, gas sensing electrodes, oxygen electrode, Biosensors.

Unit-I

The beginning of microbiology and Microbial Characteristics

Introduction to Microbiology and Microbes; History and scope of Microbiology – Hook, Antony van Leeuwenhoek and Cohn; Contribution of Pasteur and Koch. Prokaryotic cell structure, pure culture techniques; bacterial genetics: transformation, transduction and conjugation; antimicrobial resistance. Culture collection and Maintenance of cultures.

Unit-II

Microbial Taxonomy and Microbial diversity

Criteria for classification of bacteria; Bergy's manual, Cyanobacteria, acetic acid bacteria, lactic acid bacteria and Mycobacteria. Archaea: Halophiles, Methanogens and thermophiles. Viruses: general properties of virus, viral structure, sub-viral particles – viroids and prions. Eukarya: algae and fungi, general characteristics and outline classification.

Unit-III

Microbial Growth and Control

Microbial growth: Growth curve, batch and continuous culture system culture, factors affecting growth like temperature, acidity, alkalinity. Sterilization, disinfection and antisepsis: physical and chemical methods for control of microorganisms, antibiotics, Microbes and environment: Nutrient cycles (carbon and nitrogen cycle); microbial communication system; quorum sensing, prebiotics and probiotics.

Unit-IV

Beneficial and Harmful effects of Microorganism

Beneficial aspects of microbes and their metabolites in food industry, Bioremediation. Important microbial diseases of Plants caused by fungi, bacteria and viruses. Important infectious diseases of humans, caused by bacteria, protozoa and viruses - tuberculosis, malaria and AIDS. Emerging and resurgent infectious diseases.

Unit-I

Carbohydrates, Lipids and Nucleic acids

Carbohydrates: Structure of starch, glycogen and bacterial cell wall polysaccharides. Structure and biological significance of glycoproteins and proteoglycans.

Lipids: Classification, structure and functions of storage and membrane lipids- TAG, phospholipids, sphingolipids, glycolipids, isoprenoids and eicosanoids.

Nucleic acids: Structure of DNA, chemical synthesis of DNA, Isolation and characterization. structure of RNA, types and functions.

Unit-II

Proteins: Amino acids- structure and functional group properties, peptide bond, structural organization of proteins- primary, secondary, super-secondary, tertiary and quaternary, protein structures- myoglobin, collagen, keratin, immunoglobulin, Ramachandran plot, end group analysis, primary structure determination, synthesis of peptides, structure-function relationships in model proteins- Myoglobin, Haemoglobin, denaturation and renaturation of proteins- Ribonuclease A

Unit-III

Enzymology

Classification, enzyme activity, Michaelis-Menten kinetics, LB plot, inhibition - competitive, uncompetitive, non-competitive, mixed, partial, substrate inhibition, suicide inhibition, determination of K_i , active site, allosterism - ATCase, isoenzymes- LDH, catalytic strategies, co-enzymes and cofactors, multienzyme complexes- PDC.

Unit-IV

Bioenergetics

Electron transport chain and Oxidative phosphorylation: organization of respiratory chain complexes, structure and function of components, Oxidative phosphorylation. Mechanism of ATP synthesis, ATP synthase complex, proton motive force, Mitchell's hypothesis, mitochondrial permeability transition pore and its implications. Overview of Integration of metabolic pathways to bioenergetics- Glycolysis, TCA cycle, Glycogen metabolism, Pentose phosphate pathway, Gluconeogenesis, Amino acid metabolism, fatty acid metabolism, Nucleic acid metabolism

Practical-1 (Bioanalytical Techniques, Microbiology, Biochemistry)

HC

- Measurement of pH.
 - Preparation buffers and solutions.
 - Determination of pKa of amino acids.
 - Estimation of reducing sugar by DNS method.
 - Estimation of proteins by Lowry's method.
 - Estimation of proteins by Bradford's method.
 - Estimation of proteins by Bicinchonic acid method.
 - Wavelength scan of proteins and nucleic acids.
 - Ascending, descending and circular paper chromatography for separation of amino acids.
 - TLC of amino acids (1D and 2D).
 - Ultracentrifugation.
 - UV-Visible Spectrophotometry.
 - Column chromatography- gel filtration.
 - Gel electrophoresis- native and SDS-PAGE and estimation of molecular weight of proteins.
 - Demonstration of HPLC, LC-MS, XRD, NMR, Confocal and Electron microscopy.
 - Assay of acid phosphatase- Specific activity, effect of pH, determination of Km, Vmax, IC50 value.
 - Preparation of liquid and solid media for growth of microorganisms.
 - Isolation and maintenance of organisms by plating, streaking and serial dilution methods, slants and stab cultures, storage of microorganisms.
 - Isolation of pure cultures from soil and water.
 - Growth, growth curve; measurement of bacterial population by turbidometry and serial dilution methods. Effect of temperature, pH, carbon and nitrogen sources on growth.
 - Microscopic examination of bacteria, yeast and molds and study of organisms by gram stain, acid fast stain and staining for spores.
 - Assay of antibiotics and demonstration of antibiotic resistance.
 - Biochemical characterization of selected microbes.
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Softcore Papers – Semester 1

Cancer Biology (SC)

48 h

Unit-I

Cancer Biology: the basics

Introduction, historical perspective, classification, Carcinogenesis, cancer initiation, promotion and progression, Cancer cell cycles, Genomic instability, Apoptosis, Genes and proteins as players in apoptosis, DNA viruses/ cell immortalization.

Unit-II

Cancer Genes I: Oncogenes and signal transduction

Cellular proto-oncogenes, oncogene activation, Growth factors, growth factor receptors, signal transduction, Transcription, Transcription factors and cancer, Retroviral oncogenes, Tumor suppressor, Tumor suppressor gene pathways, DNA methylation, epigenetic silencing of suppressor genes.

Unit-III

Understanding Cancer as a Disease: natural history of cancer development

Free radicals, antioxidants and metabolic oxidative stress and cancer, Epidemiology of selected cancers, Gene rearrangements, detecting oncogene abnormalities in clinical specimens, Cell: cell interactions, cell adhesion, angiogenesis, invasion and metastasis, Antiangiogenic therapy of cancer.

Unit-IV

Current concepts in cancer therapy

Strategies of anticancer chemotherapy, Strategies of anticancer gene therapy/translating therapies from the laboratory to the clinic, Gene discovery in cancer research, cancer genome anatomy project, Cancer immunity and strategies of anticancer immunotherapy, stem cells and their applications in cancer therapy.

Unit-I

Introduction to Food biotechnology, Fermented foods, milk-based products, fermented vegetables, fermented meats, fish, beverages, vinegar, mould fermentation - tempeh, soysauce, rice wine. Enzymatic processing of fruit juices; DNA-based methods for food authentication, comparative methods of toxicity testing in (novel) foods, application of generic technologies in food and nutritional sciences; anti-cancer components in foods.

Unit-II

Functional foods and Biotechnology: Biochemical processing in the improvement of functional foods with targeted health benefits and increased nutrient value; Pre- and Pro- biotics, single cell protein, single cell lipids. Manipulation of fruit ripening process. Food processing, principles and practices, food ingredients and processing aids from biotechnological processes, corn sweeteners, bacterial starter cultures, cold-adapted enzymes. Food spoilage, preservation, mycotoxins in food commodities. Genetically modified foods, designer foods, detection of GM foods, Nutraceuticals, Concept of food parks.

Unit-III

Introduction to Environment, Renewable and non-renewable resources, current status of biotechnology in environment protection. Waste water management: Bioreactors for waste-water treatment, treatment of industrial effluents-dairy, distillery, paper and sugar industries. Membrane-based waste water treatment. Biotechnology & Environment, Biodiversity and its conservation, Microbial ecology.

Unit -IV

Bioremediation: Concepts and principles, bioremediation using microbes, in situ and ex situ bioremediation, biosorption and bioaccumulation of heavy metals. Phytoremediation Xenobiotics: Degradation capabilities of microorganisms with reference to toxicology, pesticides, herbicides, polyaromatic hydrocarbons.

Biostatistics

Unit I

Statistical concepts: Data structure, sampling methods, descriptive statistics - data collection, tabulation Measures of central tendency: mean, median, mode Measures of dispersion: Range, interquartile range, mean deviation, standard deviation, standard error, coefficient of variation, confidence limits.

Unit II

Types of distribution of data: Normal, Binomial, Poisson

Hypothesis testing: Z-test, t-test, ANOVA, multiple comparisons – LSD and DMRT, chi-square test; Regression and correlation; Non-parametric significance tests; Experimental designs- CRBD, RCBD, LSD, factorial; data transformation- arcsine, log, square-root. Probability

Bioinformatics

Unit III

Bioinformatics- an overview, Definition and History, Applications of Bioinformatics.

Introduction to Genomics: Genome mapping, Genome sequencing, human Genome project.

Introduction to Proteomics: Tools and techniques in proteomics.

Sequence formats. Homology and similarity.

Introduction to Data mining, NCBI, EBI, DDBJ,

Database search software: ENTREZ, SRS, Expasy.

Protein Sequence Databases, UNIPROT, Structure Database: PDB.

Sequence Analysis: definition of sequence analysis, Introduction to Sequences, alignments and Dynamic Programming; Local alignment and Global alignment (algorithm and example), Pair wise Alignment, and significance of alignment, Tools of sequence alignment, Homology sequence search, Nucleotide Sequence Analysis, Protein Sequence Analysis, Parameters of Blast, BlastN, BlastP, Interpreting Blast Results.

Unit IV

Multiple sequence analysis, scoring pattern, exhaustive and heuristic algorithms; Parameters of CLUSTAL-W and CLUSTALX for multiple sequence alignment, interpretation; Phylogenetic analysis: methods and tools.

RASMOL Display Styles- Wire Frame, Ball and Stick, Space Fill, Ribbons, Cartoons.

Drug discovery: Introduction, drug discovery technologies, virtual high-throughput *in silico* screening, Target validation EMBOSS Introduction to emboss Software package and its key features, other latest commercial softwares

Molecular Biology (HC)

48 h

Unit-I

DNA helix topology: closed and super-coiled DNA, DNA topoisomerases.

DNA replication: Semi conservative, bidirectional, semidiscontinuous replication, Enzymes in DNA replication in prokaryotes- initiation, elongation and termination. Eukaryotic DNA replication, replicons, eukaryotic DNA polymerases, role of other proteins and enzymes in replication, end-replication problem, telomeric DNA and telomerase, Replication of organelle genomes, fidelity of replication, inhibitors of replication

Unit-II

Transcription: Transcription unit, RNA polymerase in prokaryotes, bacterial promoters, mechanism of transcription- initiation, elongation and termination, eukaryotic transcription, eukaryotic RNA polymerases, eukaryotic promoters- Class I, II, III, Enhancers and silencers, transcription factors, initiation, elongation and termination of transcription, inhibitors of transcription, mRNA processing- capping, polyadenylation, splicing, rRNA and tRNA processing, structural organization of mRNA, tRNA and rRNA, nuclear export of mRNA and mRNA stability.

Genetic code: Elucidation, triplet binding assay, Wobble hypothesis.

Unit-III

Translation: Composition and ultrastructure of prokaryotic and eukaryotic ribosomes, partial reconstitution experiments, amino acid activation, amino acylation of tRNA, aminoacyl tRNA synthetases, prokaryotic and eukaryotic translation- mechanism of initiation, elongation and termination, inhibitors of translation

Protein localization: Synthesis of secretory proteins and membrane proteins, import into nucleus, mitochondria, chloroplasts and peroxisomes, post translational modifications- signal cleavage, covalent modification, Protein folding, protein degradation pathway.

Unit-IV

Regulation of gene expression in Prokaryotes: Basic control circuits, positive and negative regulation, operon concept-*lac*, *ara* and *trp* operons, catabolite repression, regulatory elements in prokaryotes, attenuation, antitermination, stringent response, regulation of gene expression in bacteriophage - lytic and lysogenic cycle.

Regulation of gene expression in Eukaryotes: Cis control elements- promoters, enhancers, Trans acting factors, DNA binding motifs of transcription factors, mechanism of regulation by transcription factors- activators and repressors, NFkB pathway, role of chromatin in regulating gene expression and gene silencing, chromatin remodeling complexes, histone acetyl transferase and deacetylase, DNA methylation and gene regulation, hormonal regulation of gene expression (peptide and steroid hormones), post- transcriptional control- alternative splicing, RNA editing, translational control- regulation of ferritin and transferrin receptor mRNA, RNA interference, gene silencing by siRNA and miRNA.

Unit-I

Cloning and Expression vectors: Plasmids, lambda vectors, M13 Phage, cosmids, phagemids, Artificial chromosome vectors-YACs, PACs and BACs, plant and animal viruses as vectors, Transposons, Expression vectors- prokaryotic (pRSET, pET), eukaryotic (pcDNA3, pCEP), Baculovirus and Pichia vector system, plant based vectors- Ti and Ri, binary and shuttle vectors, Gene cloning: genomic cloning, c-DNA cloning,

Unit-II

Gene manipulation Restriction enzymes, restriction mapping, cloning in plasmid, Phage and cosmid vectors, insertion of foreign DNA into host cells-transformation, electroporation, Transfection transient and stable, screening methods for transformants, downstream processing of recombinant proteins, affinity tags- His-tag, GST-tag, MBP-tag, Fc-tag. Construction and screening of genomic and cDNA libraries, chromosome walking, Chromosome Jumping, BAC libraries and assembly of BACs into contigs.

Unit-III

Gene analysis techniques

Hybridization techniques- Southern, Northern, South-western, Far-western, Colony hybridization, fluorescence *in situ* hybridization, molecular probes-preparation, labelling, amplification, applications, Polymerase chain reaction-Principle, primer designing, Types- RT-PCR, Realtime PCR, colony PCR, Multiplex PCR, Hot-start PCR, asymmetric PCR, Sequencing methods- chemical sequencing of DNA (Maxam and Gilberts methods and Sangers dideoxy method), automated DNA sequencing, sequencing by DE-MALDI- TOFMS, microarray. ChIP and Chip-on-chip techniques Chromogenic *in situ* hybridization, qPCR,

Unit-IV

Gene therapy, transgenics and Genome editing

Ex vivo and *in vivo* gene therapy, Vectors and other delivery systems for gene therapy, *In vitro* gene therapy, gene therapy of genetic diseases: eg. Neurological, metabolic disorders and cystic fibrosis, viruses for gene therapy- lentivirus, adenovirus. Gene targeting, knockout mice, genome editing by CRISPR-CAS

Unit-I

Immunity and nonspecific immune system: Immunity, mechanical, chemical and physiological factors, phagocytosis, humoral factors, lymphocytic cells.

Antigens and immunogenicity: The immune response, immunogenicity, molecular differences in epitope structure.

Immunoglobulins: General structure, structure and functions of specific immunoglobulins, antibody diversity, plasma cell dyscrasias.

Unit-II

The complement system: Complement, pathways of complement activation, membrane attack pathway, biological consequences of complement activation, regulatory mechanisms.

The immune response system: Exposure to an antigenic substance, the lymphoid system, cells involved in the immune response, events in the induction of the immune response, intracellular events occurring during cell maturation, phases of the humoral immune response.

Unit-III

Immune regulation: Introduction, immunosuppression, tolerance, immunopotential.

Immunization: Introduction active immunization, passive immunization, experimental immunization procedures, adverse reactions of vaccines.

Immunological techniques: *In vitro* antigen- antibody reactions, procedures for direct observation and demonstration of reactions, complex serological procedures, assays of immune competence, identification of specific allergens in type I hypersensitive reactions, detection of immune complexes, production and use of monoclonal antibodies.

Immunologic mechanisms of tissue damage: Introduction, immediate hypersensitivity (type I) reactions, cytotoxic (type II) reactions, immune complex- mediated (type III) reactions, cell-mediated (type-IV) reactions: delayed hypersensitivity and cell –mediated cytotoxicity.

Unit-IV

Auto immune diseases: General considerations, representative auto immunodisorders.

Immunodeficiency disorder: Phagocytic cell defects, B-cell deficiency disorders, T-cell deficiency disorders, secondary immunodeficiency disorders combined B-cell and T-cell deficiency disorders, secondary immunodeficiency conditions, complement deficiencies.

Transplantation immunology: Introduction, histocompatibility gene complex, clinical transplantation immunology.

Tumor immunology: Neoplasms, tumor-associated antigens, immune response to tumor antigens, immunologic factors favouring tumor growth, immunotherapy.

Practical-2 (Molecular Biology, Genetic Engineering, Immunotechnology) (HC)

- Estimation of DNA by Diphenylamine (DPA) method. •
- Estimation of RNA by orcinol method. •
- Isolation of DNA different samples: plant leaves, coconut endosperm, yeast, animal tissues. •
- Determination of purity and concentration of isolated DNA using spectrophotometer. •
- Isolation of plasmid DNA from *E. coli*. •
- Agarose gel electrophoresis of DNA. •
- Purification of DNA from gel. •
- Determination of RNase activity. •
- Isolation of RNA & analysis using Bleach Gel electrophoresis. •
- Restriction digestion of plasmid and analysis. •
- DNA ligation. •
- Transformation and screening. •
- Production of recombinant protein. •
- Polymerase chain reaction. •
- Demonstration of realtime PCR and Next generation sequencing. •
- Preparation of antigen and antibody production. •
- Purification of IgG. •
- Slide agglutination test/ Blood grouping. •
- Immunoprecipitation test- Ouchterlony double diffusion. •
- Immunoaffinity purification of IgG. •
- Immunofluorescence for localization of an antigen. •
- ELISA for quantification of an antigen. •
- Western blotting and detection. •

Softcore Papers – Semester
II Molecular Genetics (SC)

48 h

Unit-I

Genomic organization: Prokaryotes, eukaryotes, viral genome-DNA & RNA viruses extrachromosomal genome-plasmids, mitochondria and chloroplast, C-value paradox, Repetitive DNA-satellite DNAs and interspersed repeated DNAs, LINES, SINES, Alu family.

Mobile genetic elements: discovery, insertion sequence in prokaryotes, complex transposons (Tn10, Tn5, Tn9 and Tn3 as examples), mechanisms, control. Transposable elements in eukaryotes- Maize, Drosophila and humans

Unit-II

Mutation: Types, causes and detection, mutant types – lethal, conditional, biochemical, loss of function, gain of function, germinal verses somatic mutants, Molecular basis of mutations, insertional mutagenesis, site-specific mutagenesis

Recombination: Homologous and non-homologous recombination, Holliday model, site-specific recombination.

DNA Repair: Mechanism of genetic repair- direct repair, photoreactivation, excision repair, mismatch repair, post-replicative recombination repair, Repair of double-strand breaks, SOS repair.

Unit-III

Microbial genetics: Methods of genetic transfers – transformation, conjugation, transduction and sex-duction, mapping genes by interrupted mating, fine structure analysis of genes.

Gene mapping methods: Linkage maps, tetrad analysis, mapping with molecular markers, mapping by using somatic cell hybrids, development of mapping population in plants.

Quantitative genetics: Polygenic inheritance, heritability and its measurements, QTL mapping.

Unit-IV

Genes and development: **Model systems for studying development- *Drosophila*, *Caenorhabditis*, *Arabidopsis*.**

Genetic control of development in *Drosophila*: Anterior-posterior axis specification, role of maternal genes, segmentation of larval body, gap genes, pair rule genes, homeotic genes, complex gene interaction in development, sequential gene action. Floral meristems and floral development in *Arabidopsis*, ABC model.

Unit-I

Genome: Brief overview of prokaryotic and eukaryotic genome organization; extrachromosomal DNA: bacterial plasmids, mitochondria and chloroplast

Genome mapping: Genetic and physical maps; markers for genetic mapping; methods and techniques used for gene mapping, physical mapping, linkage analysis, cytogenetic techniques, FISH technique in gene mapping, somatic cell hybridization, radiation hybrid maps, *in situ* hybridization, comparative gene mapping.

Genome sequencing: Next generation sequencing, Human Genome Project, genome sequencing projects for microbes, plants and animals, accessing and retrieving genome project information from the web.

Unit-II

Comparative genomics: Identification and classification of organisms using molecular markers- 16S rRNA typing/sequencing, SNPs; use of genomes to understand evolution of eukaryotes, track emerging diseases and design new drugs; determining gene location in genome sequence.

Functional genomics: Transcriptome analysis for identification and functional annotation of gene, Contig assembly, chromosome walking and characterization of chromosomes, mining functional genes in genome, gene function- forward and reverse genetics, gene ethics, Pharmacogenomics & Personalized medicine.

Unit –III

Introduction to proteomics: Proteome and nature of proteome, Proteins - amino acids, peptides and polypeptides, separation of proteins /peptides by single and two-dimensional gel electrophoresis and detection- staining and immunoblot

Unit-IV

Structural and functional proteomics: Mass spectrometry – fundamentals, mass spectrometry ionization techniques, mass analyzers – MALDI-TOF, MS-MS, LC-MS-MS; In-gel digestion, PMF, Mass spectra analysis – search engines: Mascot, swiss-prot, protein prospector, identification, molecular weight, determination of peptide sequence, determination of post-translational modifications, peptide sequencing using tandem mass spectrometry, quantitative proteomics-iTRAQ, functional annotation of proteins, protein chips and functional proteomics; clinical and biomedical applications of proteomics

Unit I

Dynamic organization of the cell

Ultra-structure of prokaryotic and eukaryotic cells; Universal features of cells; Characteristics of cancer cells. Structure of plant cell wall, structure of cell membrane and models, functions of cell membrane intracellular organelles: Golgi apparatus; Mitochondria, chloroplast, Lysosomes Nucleus-Internal organization, Chromatin- structure and function, cellular cytoskeleton.

Unit- II

Cellular processes

Cell cycle and its regulation; cell division: mitosis, meiosis and cytokinesis: cell differentiation: stem cells, their differentiation into different cell types and organization into specialized tissues; Apoptosis.

Molecular mechanisms of membrane transport active, passive, facilitated.

Unit III

Basics of Signal Transduction

Extra-cellular matrix components, Cell junctions, Cell adhesion molecules, Hormones and their receptors, Cell surface receptors as reception of extra-cellular signals, Types of cell signaling, Growth factors- EGFR, VEGF, PDGF and their Signaling, adapter proteins required for signal transmission; signaling through G-protein coupled receptors; Second messengers in signal transduction pathways: cAMP and calcium ions (Ca²⁺), signaling through Receptor tyrosine kinases;

Unit-IV

Signal transduction pathways in animals: MAP kinase, Intracellular signaling in

Development and Disease, SAP/JNK, p38, Wnt signaling, Jak/Stat, Smad, TGF · Signaling, Cytoskeleton And Cell Signalling, MMPs And Cell Signalling, Cross talks among cytoplasmic components, NF-· B signaling from cytoplasm to nucleus. Nuclear receptors and transcription factors in signaling.

Host-parasite interaction: pathogen-induced signaling pathways in plants- ROS, Jasmonate, SA-mediated pathways.

Unit-I: Techniques in plant tissue culture

Methods in Plant Tissue culture: Concept of cellular Totipotency, Role of phytohormones in tissue culture techniques. Establishment of cultures- Nutritional requirements for in vitro cultures, Media preparation and sterilization.

Micropropagation: Propagation from shoot apical meristem, node cultures, stages of micropropagation and applications. **Germplasm preservation:** Plant germplasm storage using different methods. **Haploid Production:** Methods of androgenic haploid cultures. **Protoplast Culture and Somatic Hybridization:** Protoplast isolation, purification and culture, protoplast fusion, somatic hybridization, applications of somatic hybrids.

Unit-II: Genetic manipulation of plants

Plant transformation techniques: Agrobacterium-plant interaction, Ti plasmids, T-DNA transfer, disarmed Ti plasmid. Agrobacterium-mediated gene delivery- binary and co-integrated vectors.

Direct gene transfer methods- Particle bombardment, PEG-mediated, electroporation.

Transgenic plants: Herbicide resistance, pest resistance, plant disease resistance, improvement of nutritional quality. Biosafety regulations of transgenics.

Unit III: Applications of Plant Tissue culture

Secondary metabolite production: Major secondary metabolic pathways- Phenylpropanoid pathways, Shikimate pathway; Induction of bioactive secondary metabolites by plant tissue culture; Value addition via biotransformation; hairy root cultures for production of pharmaceuticals. Bioreactor systems for mass cultivation of plant cells, Molecular pharming: edible vaccines.

Unit-IV: Commercial product development

Micro algal biotechnology: Cyanobacteria, culture media, cultivation methods, Medicinal compound from cyanobacteria.

Single-Cell Proteins (SCP): Spirulina, Chlorella, Yeast as SCP; Production and process; Health benefits of SCP.

Agricultural products: biofertilizers and Vermiculture.

Biofuels: production of Ethanol, Methane, and their applications.

Intellectual Property Rights (IPR): IPRs and agricultural technology- implications for India. Plant Breeder's Rights. Labeling of GM crops and foods. Biodiversity, traditional knowledge, access and benefit sharing.

Animal Biotechnology (HC) 48h

Unit-I

Culture of animal cells: Advantages and limitations of tissue culture, aseptic handling, facilities required, media and cell lines. Primary culture: Isolation of mouse and chick embryos, human biopsies, methods for primary culture, nomenclature of cell lines, sub culture and propagation, immortalization of cell lines, cell line designation, selection of cell line and routine maintenance. Secondary cell culture,

Cloning and Selection: Cloning protocol, stimulation of plating efficiency, suspension cloning, isolation of clones, isolation of genetic variants, interaction with substrate, selective inhibitors.

Unit-II

Cell separation and characterization: Density based, antibody based, magnetic and fluorescence based cell sorting. Characterization of cells based in morphology, chromosome analysis, DNA content, RNA and protein, enzyme activity, antigenic markers, cytotoxicity assays, cell quantitation, cell culture contamination: monitoring and eradication, cryopreservation.

Culturing of specialized cells: Epithelial, mesenchymal, neuro ectodermal, hematopoietic gonad and tumor cells, Lymphocyte preparation, culture of amniocytes, fish cells, confocal microscopy. Stem cell culture and its applications

Organic and embryo culture: Choice of models, organ culture, histotypic culture, filter-well inserts, neuronal aggregates whole embryo culture eggs, chick and mammalian embryos.

Unit-III

Cell and Tissue engineering: Growth factors for *in situ* tissue regeneration, biomaterials intissue engineering, approaches for tissue engineering of skin, bone grafts, nerve grafts. Haemoglobin-based blood substitutes, bio artificial or biohybrid organs. Limitations and possibilities of tissue engineering, 3D bioprinting. ***In vitro* fertilization and Embryo transfer:** *In vitro* fertilization in Humans, Embryo transfer in Humans, Super ovulation and embryo transfer in farm animals e.g: Cow.

Cloning of Animals: Methods and uses. Introduction, nuclear transfer for cloning, cloning from- embryonic cells, adult and fetal cells. Cloning from short-term cultured cells: cloning of sheep, monkeys, mice, pets, goats and pigs. Cloning from long-term cultured cells: Cloning of cows from aged animals. Cloning efficiency, cloning for production of transgenic animals, gene targeting for cloned transgenic animals, cloning for conservation, human cloning: ethical issues and risks.

Unit-IV

Transfection methods and transgenic animals: Gene transfer, transfection of fertilized eggs or embryos, unfertilized eggs, cultured mammalian cells, targeted gene transfer. Transgenic animals and applications: mice and other animals, sheep, pigs, goats, cows and fish.

The legal and socio-economic impact of biotechnology at national and international levels, public awareness.

Biosafety regulations- guidelines for research in transgenic animals, public awareness of the processes of producing transgenic organisms.

Unit I

Basic principles: Isolation, screening and maintenance of industrially important microbes; effect of nutrients, temperature, pH for the growth of industrially important microorganisms; strain improvement for increased yield.

Batch and continuous fermenters: types of fermenters, chemostat, turbidostat, upstream processing; media formulation and optimization; sterilization; aeration, agitation, pH.

Unit II

Downstream processing:

Separation of insoluble products – separation of cells and foam; filtration (plate filters, rotary vacuum filter), centrifugation (continuous, basket and bowl centrifuge), Stokes law, sedimentation, flocculation; cell disruption (mechanical and non-mechanical methods); chromatographic techniques, drying (spray, drum, freeze driers); storage and packaging.

Unit III

Microbial products: Microbial production and application of vitamins, enzymes, organic acids (acetic, citric, gluconic, itaconic, lactic,), amino acids (glutamic acid, lysine, tryptophan), polymers (polysaccharides – xanthan, curdlan, dextran, pullulan,), antibiotics, ethanol, biosurfactants.

Unit IV

Bioprocess in agro-industry: Isolation and screening of bioagents for the production of biofertilizers, biopesticides and plant growth promotion; mass cultivation, formulation and storage life; Bioprocess in sustainable agriculture (organic matter recycling, composting, Jeevamrutha).

Practical-3 (Plant and Animal Biotechnology, Bioprocess Technology) (HC)

Plant Biotechnology

- Preparation of plant tissue culture media •
- Callus induction •
- Induction of somatic embryogenesis •
- Establishment of cell suspension cultures for plant secondary metabolite production •
- Encapsulation of somatic embryos and production of synthetic seeds •
- Organ cultures: Shoot tip, nodal, anther and leaf cultures •
- Micropropagation technique – banana •
- Protoplast isolation technique •
- Secondary metabolite estimations: Colorimetry and TLC methods •

Animal Biotechnology

- Preparation of media, culture and maintenance of cell lines, trypsinization
- Culture of transformed cells
- MTT assay for cytotoxicity
- Lymphocyte preparation

Bioprocess Technology

- Immobilization of yeast by calcium alginate gel entrapment and assay for enzymes- invertase and catalase
 - Screening of antibiotic producing microorganisms
 - Study of alcohol fermentation- alcohol from different substrates-estimation of alcohol content
 - Bioassay methods- Vitamins and amino acids
 - Analysis of microbial quality of foods
 - Study of fermenter (demonstration)
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Softcore Papers – Semester III

Molecular Diagnostics (SC) 48h

Unit-I:

Introduction to Plant molecular diagnostics, Conventional diagnostic techniques for the detection of plant pathogens – fungi, bacteria and viruses, seed-borne pathogens, Koch rules, Direct detection and identification of pathogenic organisms. Culture based and GOTs, Indirect detection of pathogenic organisms, Serology based detection – IF, ODD, ELISA, DIBA, ISEM. Automated identification methods.

Unit-II

Genome resolution, detection and analysis: Different types of PCR: Real-time; Multiplex; FISH; RFLP; DGGE; SSCP; Nucleic acid sequencing: new generations of automated sequencers; Microarray chips; EST; SAGE; microarray data normalization & analysis; molecular markers: 16S rRNA typing; MALDITOF-MS; Metabolite profile for biomarker detection the tissues in various disorders by making using LCMS & NMR technological platforms.

Unit-III

Background and scope of molecular diagnostics, Current and emerging technologies, Nonamplified Probe-Based Assays, Cytogenetics and FISH, PCR and Other Amplification Technologies, Microarrays, DNA Methylation Assays, Infectious Disease Diagnostics: Human Immunodeficiency Virus, Hepatitis C Virus, Hepatitis B Virus, Molecular Tests for Blood Screening, Chlamydia and Gonorrhoea, Human Papillomavirus and Cervical Cancer, Methicillin-Resistant Staphylococcus aureus, Genetic Testing: Tests for Inherited Disorders and Prenatal Diagnostics: Cystic Fibrosis, Prenatal Diagnosis, Ashkenazi Jewish Genetic Panel, Predicting Risk of Disease, Risk of Venous Thromboembolism

Unit -IV

Molecular Oncology Tests, Analysis of the Expression of Multiple Genes and Cancer Prognosis, Analysis of Lymph Nodes to Detect Metastasis of Breast Cancer, Screening for Colorectal Cancer: Stool-Based DNA Screening, Leukemias and Lymphomas, DNA Methylation Tests and Cancer, Predicting Risk of Developing Cancer. Personalized Medicine: Pharmacogenomics and Companion Diagnostics, Cytochrome P450 and Drug Metabolism, Targeted Cancer Therapies and Companion Diagnostics Tests, Testing for HER2/neu Overexpression in Breast Cancer, Testing for Epidermal Growth Factor Receptor (EGFR), UGT1A1 Genetic Variants, Pharmacogenetics and Response to Antiretroviral Therapy, Thiopurine Methyltransferase and Metabolism of Thiopurine Drugs

Unit I

Prospects of Natural Products research in the 21st Century: Introduction, use of natural products in traditional medicines, Marine natural products, Use of herbal remedies and the potential of drug development from natural products and novel drug templates: paclitaxel, podophyllotoxin, artemisinin etc. Recent development in the research on naturally occurring polyphenolic compounds: - Introduction, biosynthetic pathway, isolation and characterization, biological and pharmacological activities of different class of phytoconstituents - alkaloids, flavonoids, terpenoids, glycosides, steroids, saponins, (Antioxidant activity, cyto-toxic activity, anticancer and anti-microbial activity etc). aid design of clinical studies.

Unit II

Natural product drug discovery from different sources (marine, microbial, mineral etc): Introduction, recent developments, applications. Extraction and Isolation techniques: Introduction, Principle and Applications of different extraction & isolation methods viz Soxhlet extraction, microwave extraction, supercritical fluid extraction, solid phase extraction, Column chromatography, Flash chromatography.

Unit III

Target identification and molecular modeling: Identification of target or drug leads associated with a particular disease by different techniques including combinations of molecular modeling, combinatorial libraries and high-throughput screening (HTS); Use of bioinformatics and data processing in identification of lead compounds; Rational drug design Modelling drug/receptor interactions with the emphasis on molecular mechanisms, molecular dynamics simulations and homology modelling; Conformational sampling, macromolecular folding, structural bioinformatics, receptor-based and ligand-based design and docking methods, in silico screening of libraries, semi-empirical and ab-initio methods, QSAR methods, molecular diversity, design of combinatorial libraries of drug-like molecules macromolecular and chemical databases.

Unit IV

Lead optimization: Identification of relevant groups on a molecule that interact with a receptor and are responsible for biological activity; Understanding structure activity relationship; Structure modification to increase potency and therapeutic index; Concept of quantitative drug design using Quantitative structure–activity relationship models (QSAR models); Bioanalytical assay development in support of in vitro and in vivo studies (LC/MS/MS, GC/MS and ELISA). Preclinical development: Principles of drug absorption, drug metabolism and distribution - intestinal absorption, metabolic stability, drug-drug interactions, plasma protein binding assays, metabolite profile studies, Principles of toxicology, Experimental design for preclinical and clinical PK/PD/TK studies, Selection of animal model; Regulatory guidelines for preclinical PK/PD/TK studies; Scope of GLP, SOP for conduct of clinical & non clinical testing, control on animal house, report preparation and documentation. Integration of nonclinical and preclinical data tool.

Unit I

Introduction and Fundamentals of nanobiotechnology Concepts, historical perspective; Nanoscale materials: Definition and properties; Different formats of nanomaterial and applications; Cellular nanostructure; nanopores; Biomolecular motors; Bio-inspired Nanostructures, Quantum dots. Synthesis and characterization of different nanomaterials: Synthesis of nanomaterials from plant, microbial and animal cell sources. Characterization of nanomaterials using Optical Microscopy, Scanning Electron Microscopy, Transmission Electron Microscopy, Atomic Force Microscopy, Scanning Tunneling Microscopy, Optical Absorption and Emission Spectroscopy, Thermogravimetric Analysis, Differential Scanning Calorimetry, Thermomechanical Analysis, X-Ray, neutron diffraction. Applications of nanobiotechnology in Plant and animal cell cultures, stem cell culture and artificial organ synthesis. delivery of fertilizers, pesticides (nanocides); Nanoremediation.

Unit II

Nano-particles Concepts of Nanoparticles: Basic structure of Nanoparticles- Kinetics in nano-structured Materials- Zero dimensional, size and shape of nanoparticles; one-dimensional and twodimensional nanostructures; clusters of metals and semiconductors, bionano-particles. Bionanocomposites: Nano-particles and Microorganisms; Microbial Synthesis of Nano- materials; Biological methods for synthesis of nano-emulsions using bacteria, fungi and Actinomycetes; Plant-based nanoparticle synthesis; Nano-composite biomaterials – Fibres, devices and structures, Nano Bio-systems.

Unit III

Applications of Nanobiotechnology Applications of Nanomedicine: Nanotechnology in diagnostic applications, materials used in Diagnostics and Therapeutics. Nanomaterials for catalysis, development and characterization of nanobiocatalysts, application of nano-scaffolds in synthesis, applications of nano-biocatalysis in the production of drugs and drug intermediates. Nano-films: Thin films; Colloidal nanostructures; Self-assembly, Nanovesicles; Nanospheres; nanocapsules and their characterization. Nanoparticles for drug delivery: Strategies for cellular internalization and long circulation, strategies for enhanced permeation through various anatomical barriers. Nanoparticles for diagnostics and imaging: Concepts of smart stimuli responsive nanoparticles, implications in cancer therapy, nanodevices for biosensor development. Applications in Agriculture: Biogenic nanomaterials and their role in soil, water quality and plant protection; Smart nanoscale systems for targeted

Unit IV

Sustainable bionanotechnology: Application of industrial ecology to nanotechnology, Fate of nanomaterials in environment, environmental life cycle of nano-materials, environmental and health impacts of nano materials, Nano-materials in future - implications. Toxicity and safety of nanomaterials: Introduction to Safety of nanomaterials; Concept of Nanotoxicology – Models and assays for nanotoxicity assessment; Laboratory rodent studies. Ecotoxicologic studies: Methodology for Nanotoxicology - toxicity testing; Mechanism of nano-size particle toxicity; Reactive oxygen species mechanisms of NSP toxicity; Interactions between nanoparticles and living organisms.

Softcore Papers – Semester IV

Stem Cells And Regenerative Medicine (SC)

48 hrs

Unit I:

Introduction to Stem Cells Overview of basic and translational research of stem cells. Differentiation in early development, Preimplantation development; From implantation to gastrulation. Pluripotent stem cells I: Rodent embryonic stem cells – Origin, properties, self-renewal pathways, application. Human embryonic stem cells- Derivation and maintenance, selfrenewal pathways. Induced pluripotent stem cells- Generation, Characterization, Induced pluripotency-the underlying mechanism. Primordial and embryonic germ cells- Origin, Properties, Derivation and maintenance. Stem cells: Molecular and cellular basis of organ development

Unit II

Tissue engineering principles and perspectives; Limitations and hurdles of using embryonic stem cells in tissue engineering; Amniotic fluid and amniocentesis; Isolation and characterization of amniotic fluid-derived stem cells. New technologies for genetic modification in stem cells, CRISPR/Cas9, TALENs/ZFN. Neurogenesis and neural stem cells I- Establishment of neural tissue, Molecular basis of neural induction. Neurogenesis and neural stem cells II- Neural stem cells in brain; Pluripotent stem cell-derived neural stem cells Hematopoietic stem cells- Embryonic hematopoiesis; Hematopoietic stem cell niche; Embryonic stem cell-derived Hematopoietic stem cells. Cord blood hematopoietic stem cells, Cord blood transplantation; Characteristics, Genomics and proteomics of cord blood stem cells

Unit III

Stem cells in retina and inner ear- Sources and Properties Skin organization, Skin stem cells, bulge as a residence of skin stem cells, Cell signaling in skin stem cells. Skeletal muscle stem cells- Sources, Intrinsic and extrinsic regulation Stem cells in kidney-Anatomy of kidney development, Sources and characterization of kidney stem cells. Stem cells in liver, pancreas and intestine- Organization of adult liver and pancreas, Liver/Pancreatic stem cells, Intestinal stem cells. iPSCs for disease modeling; Models of neurological diseases, hematopoietic disorders, cardiovascular conditions, metabolic disorders. Mesenchymal stem cells- Location, isolation and culture; tissueengineering

Unit IV:

Therapeutic uses of stem cells Stem cells to treat diabetes and liver disease, β -cell replacement therapy; Sources of insulinproducing cells; Hepatocyte transplantation; Challenges and future directions Cancer stem cell theory – Isolation and characterization of cancer stem cells; Implications for cancer treatment: Stem cells to treat heart disease, Distribution of stem cells in heart; Preclinical studies. Orthopedic applications of stem cells, Biology of musculoskeletal tissues; Tissue engineering strategies for bone and cartilage defects. Neural stem cells for central nervous system repair, Therapeutic potential of neural stem cells; Cell replacement using neural stem cells. Stem cells for the treatment of muscular dystrophy, Cellular environment of a dystrophic muscle; Myogenic stem cells from embryonic stem cells and inducible pluripotent stem cells; Current stem cell-based therapeutic approaches. Regeneration of epidermis, Epidermal stem cells; Stem cells in burned and skin ulcers Regulatory aspects for stem cell research; Regulation of use of human embryonic stem cells

Molecular Plant Pathology (SC)

48 h

Unit I

The fundamentals of plant pathology: The concept of plant disease, the causal agents, the significance of plant diseases, the control of plant diseases. Fungal diseases: establishing infection – dispersal spores, finding a suitable host, spore attachment, germination process, penetration, germ-tube elongation, induction appressoria, cell-wall degrading enzymes. Development of disease – Basic concepts of necrotrophy and biotrophy, host barriers, the role of toxins and enzymes, biotrophy.

Unit II

Bacterial and viral diseases: communication between bacteria, plant penetration, attachment, stimulation gene expression, cell wall degrading enzymes, toxins, hormones, extracellular polysaccharides, determinants of host specificity. Plant viruses: Structure and replication, infection, types of viruses, viroids.

Unit III

Genetics of plant diseases and resistance: Genes and diseases, Mechanism of variability, stages of variation in pathogens, Types of plant disease resistance to pathogens. Defence mechanism of plants, Pre-existing, structural, chemical and induced biochemical defences. Resistance genes: Gene-for-gene resistance, features of cloned resistance genes. MAP kinases, ion fluxes and calcium homeostasis, The oxidative burst, Nitric oxide, (p)ppGpp signaling,

Unit IV

Application of molecular biology to conventional disease control strategies: Breeding for resistance, the use of tissue culture in plant breeding, marker-assisted breeding, identification of novel resistance gene specificities, the use of chemicals for disease control, biological control-PGPR and PGPF. Transgenic approaches for crop protection- Bt cotton and brinjal.

Unit I

Innovation and entrepreneurship in bio-business Introduction and scope in Bio-entrepreneurship, Types of bio-industries and competitive dynamics between the sub-industries of the bio-sector (e.g. pharmaceuticals vs. Industrial biotech), Strategy and operations of bio-sector firms: Factors shaping opportunities for innovation and entrepreneurship in bio-sectors, and the business implications of those opportunities, Alternatives faced by emerging bio-firms and the relevant tools for strategic decision, Entrepreneurship development programs of public and private agencies (MSME, DBT, BIRAC, Make In India), strategic dimensions of patenting & commercialization strategies.

Unit II

Bio markets - business strategy and marketing Negotiating the road from lab to the market (strategies and processes of negotiation with financiers, government and regulatory authorities), Pricing strategy, Challenges in marketing in bio business (market conditions & segments; developing distribution channels, the nature, analysis and management of customer needs), Basic contract principles, different types of agreement and contract terms typically found in joint venture and development agreements, Dispute resolution skills. Overview of Research Methodology & Project Proposal writing.

Unit III

Finance and accounting Business plan preparation including statutory and legal requirements, Business feasibility study, financial management issues of procurement of capital and management of costs, Collaborations & partnership, Information technology.

Unit IV

Technology management Technology – assessment, development & upgradation, Managing technology transfer, Quality control & transfer of foreign technologies, Knowledge centers and Technology transfer agencies, Understanding of regulatory compliances and procedures (CDSCO, NBA, GCP, GLA, GMP).

Unit-I:

Introduction Seed Biology: Floral biology, mode of reproduction; Embryogenesis and seed development; Seed structure of monocots and dicots; Chemical composition of seeds; Orthodox and recalcitrant seeds, seed dormancy; Apomixis, parthenocarpy, polyembryony; Somatic embryogenesis and synthetic seeds. Development of Seed Industry: Agricultural situation in India; impact of green revolution; cropping systems; International cooperation – ISTA, OECD, UPOV, AOSA, APSA, CGIAR and other organizations. Seed Production: Introduction to crop breeding methods; Variety testing, release and certification; Different classes of seeds and their maintenance; Seed production requirements and planning; Male sterility; Clonal propagation; Transgenic seeds. Disease tolerance screening. Seed drying, processing, storage and marketing: Seed drying principles and methods; Seed treatment, safe storage seeds and marketing strategies. , factors influencing mycotoxin production, harmful effects, detection.. Gene targets and primer designing.

Unit-II:

Seed Quality Control Importance of seed quality: Seed legislation - Seed act 1965, seed rules 1969 and new seed act 2004. Seed certification - History, concept, organization, phases and seed certification standards; Field inspection principles and methods; Determination of seed quality - seed sampling, physical purity, moisture, germination, genetic purity; Seed certification agencies; Testing of transgenic seeds.

Unit-III:

Seed Health Importance: Designated plant diseases, tolerance, seed health and trade, Pest-free areas (PFA), Pest Risk Analysis (PRA). Significance of seed health - important seed borne diseases of cereals, pulses, oil seeds, fiber and vegetable crops; Mechanism of seed transmission and disease cycle. Management of seed-borne diseases: Quarantine and phytosanitary certificates, Physical and chemical control, biological control, cross protection. Storage fungi and insects: Causes and indices of seed deterioration during storage, fumigation. Mycotoxins – Important mycotoxins

Unit-IV:

Diagnostics - Seed health testing procedures for Fungi – symptoms, dry seed examination, incubation tests, embryo extraction technique, seedling symptom test; Bacteria – symptoms, colony appearance, liquid assay, selective and semi-selective media, staining techniques, biochemical & physiological tests, pathogenicity tests, immune-fluorescent technique, Biolog; Viruses – symptoms, seed examination, growing-on test, indicator plant test, electron microscopy, ISEM, ELISA, DIBA, IC-RT-PCR; Nematodes – Extraction and identification. Application of serological methods – monoclonal and polyclonal antibodies, conventional serological techniques – precipitin tests, agglutination tests, ELISA, DIBA, and nucleic acid based techniques; Multiplex ELISA and PCR, Application of Real Time (RT)-PCR; FTA technology. Sequence databases of seed-borne pathogens

Open elective

Biotechnology and its Applications 48 h

Unit I

Introduction to biotechnology. Principles of biotechnology, classification. Recombinant DNA Technology Introduction, outline of genetic engineering procedure, restriction endonucleases, cloning & expression vectors- plasmids, cloning in plasmid, transformation and detection of transformants- lacZ, genomic and cDNA libraries, gene analysis techniques- hybridization: Southern, Northern, Western, in situ, Polymerase chain reaction.

Unit II

Microbial and food and environmental Biotechnology Basics of fermentation technology: Types of microbial culture- batch, continuous and fedbatch. Microbial production: Use of microbes in production of vitamins, enzymes, organic acids, amino acids, polysaccharides, flavors, sweeteners, proteins and antibiotics. Fermented food products- yogurt, cheese, tempeh, sauerkraut; beverages- wine and beer. Pre- and Pro-biotics, single cell proteins, Genetically modified foods, designer foods. Current status of biotechnology in environment. Bioconservation, biofuels, gasohol, biogas. Bioremediation: Concepts and principles, bioremediation using microbes, in situ and ex situ bioremediation, biosorption and bioaccumulation of heavy metals.

Unit III

Plant Biotechnology Landmarks in Plant tissue culture. Types of cultures- embryo, organ, callus and cell cultures, Somatic embryogenesis, Haploid Production, Androgenesis, Protoplast culture and somatic hybridization. Micropropagation- Methods and stages, applications. Synthetic seeds, somaclonal variation. Production of secondary metabolites by plant cells, Biotransformation. Plant transformation techniques: Direct and indirect methods of gene transfer in plants. Transgenic plants and crop improvement- herbicide tolerance, disease resistance, abiotic stress tolerance, delayed ripening, improvement of nutritional quality, molecular pharming.

Unit IV

Animal Biotechnology Basics of animal cell culture techniques, cell lines, physical conditions for culturing animal cells, equipments required, scale-up of culture methods. Application of animal cell culture- Hybridomas, production of therapeutic antibodies, stem cell technology, cell and tissue engineering. Genetic engineering of animals: Methods for gene transfer in animals, microinjection, nuclear transplantation, retrovirus-mediated gene transfer, gene knockdown techniques. Transgenic- animals- sheep, pigs, cattle, chickens; applications of transgenic animals.

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